

## Quantitative Tests for Preliminary Phytochemical Analysis

Priyanka Panth<sup>1</sup>, Dr. Mohammad Hashim Mansoori<sup>2</sup>

Department of Pharmaceutical Chemistry & M.S. College of Pharmacy,  
Email: panthpriyanka29@gmail.com

Department of Pharmaceutical Chemistry & M.S. College of Pharmacy,  
Email: hashim.m@mscollegeofpharmacy.in

### Abstract:

Medicinal plants have been used in the treatment of various diseases as they possess potential pharmacological activities including antineoplastic, antimicrobial, antioxidant, anti-inflammatory, analgesic, anti-diabetic, anti-hypertensive, anti-diarrheal and other activities. Phytoconstituents individually or in combination, determine the therapeutic value of a medicinal plant. Alkaloids, flavonoids, phenolics, tannins, saponins, steroids, glycosides, terpenes etc. are some of the important phytochemicals with diverse biological activities. The pharmacological activity of a plant can be predicted by the identification of the phytochemicals. Currently, phytochemicals are determined by various modern techniques, but conventional qualitative tests are still popular for the preliminary phytochemical screening of plants.

**Keywords:** Herbal Medicinal Plants, Phytochemical Analysis, Qualitative Tests, Reagent Preparation

### INTRODUCTION

Phytochemicals are chemicals of plant origin. Phytochemicals (from Greek: *phyton* = plant) are chemical compounds naturally present in the plants attributing to positive or negative health effects<sup>[1]</sup>. Medicinal plants used in different diseases and ailments are the richest bio reservoirs of various phytochemicals. The medicinal properties of the plants are determined by the phytochemical constituents<sup>[2]</sup>. Some of the important phytochemicals include alkaloids, flavonoids, phenolics, tannins, saponins, steroids, glycosides, terpenes, etc. which are distributed in various parts of the plants<sup>[3]</sup>. Nature is a unique source of structures of high phytochemical diversity representing phenolics (45%), terpenoids and steroids (27%) and alkaloids (18%) as major groups of phytochemicals<sup>[4]</sup>. Although, these compounds seem to be non-essential to the plant producing them, they play a vital role in survival by mediation of ecological interactions with competitors, protect them from diseases, pollution, stress, and UV rays and also contribute for colour, aroma and flavour with respect to the plant. The metabolites produced by the plants to protect themselves against biotic and abiotic stresses have turned into medicines that people can use to treat various diseases<sup>[5,6]</sup>.

Phytochemicals can be separated from the plant material by various extraction techniques. The most commonly used conventional methods include maceration, percolation, infusion, digestion, decoction, hot continuous extraction (Soxhlet extraction) etc., recently, eco-friendly techniques such as Ultrasound-Assisted Extraction (UAE), Microwave-Assisted Extraction (MAE), Supercritical Fluid Extractions (SFE) and Accelerated Solvent Extraction (ASE) have also been introduced<sup>[10,11]</sup>.

Different types of solvents viz. water, ethanol, methanol, acetone, ether, benzene, chloroform etc. are used in the extraction process<sup>[12]</sup>. Extraction of phytochemicals from the plant materials is affected by pre-extraction factors (plant part used, its origin and particle size, moisture content, method of drying, degree of processing etc.) and extraction-related factors (extraction method adopted, solvent chosen, solvent to sample ratio, pH and temperature of the solvent, and length of extraction)<sup>[10,12]</sup>.

Previously, the plant parts were directly used as such for the treatment, but now-a-days, the active principles are identified and isolated in pure form and also synthetically produced with the help of advance techniques<sup>[6]</sup>. In developing new synthetic drugs, the chemical structures derived from these phytoconstituents can be used as models<sup>[7]</sup>. Identification of phytoconstituents in the plant material helps to predict that plant's potential pharmacological activity, characterization and evaluation of evidence to support those plants' therapeutic claims, and to support the therapeutic claims of those plants against various ailments<sup>[12]</sup>. Advanced techniques like Gas Chromatography (GC), Liquid Chromatography (LC), High-Performance Liquid Chromatography (HPLC), High-Performance Thin Layer Chromatography (HPTLC) etc. are very helpful for detection of phytoconstituents both qualitatively as well as quantitatively<sup>[1]</sup>. However, when these techniques are unavailable or unaffordable, the conventional phytochemical tests which are economic, easy and require fewer resources, remain the good choice for preliminary phytochemical screening<sup>[2]</sup>. The present communication deals with the collection and compilation of maximum possible qualitative phytochemical tests from various published literatures.

## **PREPARATION OF REAGENT FOR PHYTOCHEMICAL ANALYSIS:**

### **1. Dragendroff's Reagent:**

**Stock Solution:** 5.2gm Bismuth Carbonate + 4gm Sodium Iodide + 50ml Glacial Acetic acid, boiled for few min., after 12hr. precipitated Sodium Acetate crystals are filtered by sintered glass funnel; 40ml filtrate + 160ml Ethyl Acetate + 1ml distilled water, (**stored in Amber – Colored glass bottle**).

**Working Solution:** 10ml stock solution + 20ml Acetic acid + distilled water to make final volume 100ml.

### **2. Hager's Reagent:** Saturated aqueous solution of Picric acid

### **3. Mayer's Reagent:**

**Solution A:** 1.36gm Mercuric Chloride + 60ml Distilled water

**Solution B:** 5gm Potassium Iodide + 10ml Distilled water

**Working Solution:** Solution A + Solution B + Distilled water to make final volume 100ml

### **4. Wagner's Reagent:** 1.27gm iodine + 2gm Potassium Iodide + Distilled water to make final volume 100ml

### **5. Barfoed's Reagent:** 30.5gm Copper Acetate + 1.8ml Glacial Acetic acid

### **6. Seliwanoff's Reagent:** 0.01gm Resorcinol + 6.6ml Concentrated HCL + Distilled water to make final volume 20ml

### **7. Benedict's Reagent:**

**Solution A:** 173gm Sodium Citrate + 100gm Sodium Carbonate + 800ml water, dissolve and boil to make solution clear

**Solution B:** 17.3gm of Copper Sulphate Dissolve in 100ml distilled water

**Working Solution:** Mix solution A and Solution B

#### 8. Fehling Solution:

**Solution A:** 34gm Copper Sulphate + Distilled water to make final volume 100ml

**Solution B:** 173gm Potassium Sodium Tartrate + 50gm NaOH + Distilled water to make 100ml

#### 9. Baljet's Reagent:

95ml 1% Picric Acid + 5ml 10% NaOH

#### 10. Millon's Reagent:

1gm Mercury + 9ml Fuming Nitric Acid + equal amount of distilled water (after completion reaction)

### QUALITATIVE TESTS FOR PHYTOCHEMICAL ANALYSIS:

#### Detection of Quinones:

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Alcoholic KOH test	1 mL plant extract + a few mL alcoholic potassium hydroxide	Red to blue colour	[21, 26]
2	Conc. HCl test	Plant extract + conc. HCl	A green colour	[17]
3	Sulphuric acid test	10 mg extract + dissolved in isopropyl alcohol + a drop of conc. H <sub>2</sub> SO <sub>4</sub>	A red colour	[32]

#### Detection of Reducing Sugar:

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Benedict's test	0.5 mL filtrate <sup>b</sup> + 0.5 mL Benedict's reagent + Boiled for 2 min.	Green/yellow/red colour	[7, 21]
2	Fehling's test	1 mL each of Fehling's solution A & B + 1 mL filtrate <sup>b</sup> + boiled in water bath	A red precipitate	[7, 21]

#### Detection of Proteins and Amino acids:

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Biuret test	2 mL filtrate + 1 drop of 2% copper sulphate sol. + 1 mL of 95% ethanol + KOH pellets	A pink-coloured sol. (in an ethanolic layer)	[1, 7]
2	Millon's test	2 mL filtrate + a few drops of Millon's reagent	A white precipitate	[1, 7]

3	Ninhydrin test	2mL filtrate + 2 drops of Ninhydrin solution (10mg ninhydrin + 200mL acetone)	A purple coloured sol. { Amino acids }	[1, 7]
4	Xanthoproteic test	Plant extract + Few drops of conc. Nitric acid	A yellow-coloured sol.	[1, 12]

**Detection of Alkaloids:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Dragendorff's Test	Few ml Filtrate <sup>a</sup> + 1-2 ml Dragendorff's reagents	A reddish-brown precipitate	[1, 21]
2	Hager's test	Few mL filtrate <sup>a</sup> + 1-2 mL Hager's reagents	A creamy white precipitate	[1, 7]
3	Mayer's/Bertrand's/Valser's test	Few mL filtrate <sup>a</sup> + 1-2 drops of Mayer's reagent (Along the sides of test tube)	A creamy white/yellow precipitate	[7, 12, 13]
4	Wagner's test	Few mL filtrate <sup>a</sup> + 1-2 drops of Wagner's reagent (Along the sides of test tube)	A brown/reddish precipitate	[7, 21]
5	Picric acid test	Few mL filtrate <sup>a</sup> + 3-4 drops of 2% picric acid solution	A orange colour	[14, 15]
6	Iodine Test	3 mL extract solution + few drops of iodine solution	A blue colour, which disappears on boiling and reappears on cooling	[16, 17]
7	Bouchardat's test	6 mL plant extract, evaporated completely + 6 mL ethanol (@ 60°C) + few drops of Bouchardat's reagent (dilute iodine solution)	A reddish brown colour	[18]
8	Tannic acid test	Acidified extract + 10% tannic acid solution	A buff colour precipitate	[30, 38]

**Detection of Carbohydrate:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Barfoed's test	1 mL filtrate <sup>b</sup> + 1 mL Barfoed's reagent + Heated for 2 min.	A red precipitate { monosaccharides }	[7, 19]
2	Molish's test	2 mL filtrate <sup>b</sup> + 2 drops of alcoholic $\alpha$ -naphthol + 1 mL conc. H <sub>2</sub> SO <sub>4</sub> (along the sides of test tube)	A violet ring	[7, 21]
3	Seliwanoff's Test	1 mL extract solution + 3 mL seliwanoff's reagent + heated on water bath for 1 min.	A rose red colour { ketoses }	[17, 19]

4	Resorcinol test	2mLaq.extractsolution+fewcrystalsofresorcinol +equal volumeof conc.HCl +heated	Arosecolour {ketones}	[13, 9]
5	Testforpentos	2mLconc.HCl+ littleamountofphloroglucinol+equalamountof aqueousextractsolution+ heatedoverflame	A redcolour	[13]
6	Testforstarch	Aqueousextract+ 5mL5%KOH solution	Acinarycolouration	[20]

**Detection of Glycosides:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Bortrager's test	2mLfiltratedhydrolysate <sup>c</sup> +3mLChloroform +shakenwell +chloroformlayer isseparated+10% Ammoniasolution	A pink colouredsolution	[7]
2	Modified Bortrager's test	Plantextract+ferric chloridesolution+boil for5min. + cooled + equal volome of benzene + benzene layer isseparated+ Ammoniasolution	Arose-pink to blood redcolouredsolution	[12, 33]
3	Legal's test	Dissolve 50gmplantextractinpyridine+ Sodiumnitroprusside +10%Sodiumhydroxide	A pink colouredsolution	[7, 12]
4	10% NaOH test	1mL dil. H <sub>2</sub> SO <sub>4</sub> + 0.2mL extract + boiled for 15min.+allowedcooling+neutralize with10% NaOH+0.2mL Fehling's solution A&B	A brickred precipitate	[21]
5	Aqueous NaOH test	Alcoholic extract+dissolvedin 1mLofwater +few drops of aqueousNaOHsolution	A yellow colour	[22]
6	Concentrate H <sub>2</sub> SO <sub>4</sub> test	5mplantextract+2mLglacial aceticacid +adrop of5% FeCl <sub>3</sub> +conc.H <sub>2</sub> SO <sub>4</sub>	A brown ring	[3]
7	Raymond's test	Extractsolution+dinitrobenzeneinhotmethanolicalkali	A violet colour	[38]

**Detection of Cardiac Glycosides:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
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1	Keller-Killanitest	1mLfiltrate+1.5mLglacialaceticacid+ 1 drop of 5% ferric chloride + conc. H <sub>2</sub> SO <sub>4</sub> (along the sideof testtube)	A blue coloured solution(inacetic acidlayer)	[21, 38]
2	Kedee'stest	4mLextractevaporatedtodryness+1-2mLmethanol+1-2mL alcoholic KOH + 3-4 dropsof 1% alcoholic 3,5-dinitrobenzene+heated	Adisappearing violetcolour {Cardenolides}	[22]
3	TestforCardenolides	Extract+pyridine+Sodiumnitroprusside +20%NaOH	Aredcolour, fades to brownishyellow	[20]
4	Brominewater test	Plantextract+fewmL ofbrominewater	Ayellowprecipitate	[30]
5	Baljettest	2mLextract+adropofBaljet'sreagent	Ayellow-orangecolour	[39, 40]

**Detection of Flavonoids:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Alkaline reagent test	1mL extract+2mL of 2% NaOH solution (+few drops dil. HCl)	An intense yellow colour, becomes colourless on addition of diluted acid	[20, 21, 23]
2	Hager's test	Plant extract+10% ammonium hydroxide sol.	A yellow fluorescence	[7]
3	Lead acetate test	1mL plant extract+few drops of 10% lead acetate solution	A yellow precipitate	[1, 21, 12]
4	Shinoda's test/Mg-hydrochloride reduction test	Plant extract is dissolved in 5mL alcohol+Fragments of magnesium ribbon+few drops of conc. HCl	A pink to crimson coloured solution {flavonol glycosides}	[7, 38]
5	Shibata's reaction/ Cyanide test	1gm Aq. extract+dissolved in 1-2mL 50% methanol by heating+metal magnesium+5-6 drops of conc. HCl	A red colour {flavonols}, orange colour {flavones}	[13, 22]
6	Ferric chloride test	Extract aqueous solution+few drops 10% ferric chloride solution	A green precipitate	[20]
7	Pew's test	Few mL aqueous extract solution+0.1g metallic zinc+ 8mL conc. H <sub>2</sub> SO <sub>4</sub>	A red colour {flavonols}	[13]
8	Zinc-hydrochloride	Plant extract+ pinch of zinc dust+ conc. HCl along the side	Magenta colour	[24, 25]

	reduction test	of test tube		
9	Ammonia test	Filtrate+5mL dil. Ammonia solution+conc. H <sub>2</sub> SO <sub>4</sub>	A yellow colour	[26]

**Detection of Phenolic Compounds:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Iodine test	1mL extract+few drops of dil. Iodine sol.	A transient red colour	[21]
2	Ferric chloride test	Extract aqueous solution+few drops 5% ferric chloride sol.	Dark green/bluish black colour	[7, 12]
3	Gelatin test	Plant extract is dissolved in 5mL distilled water+ 1% gelatin solution+10% NaCl	A white precipitate	[7]
4	Lead acetate test	Plant extract is dissolved in 5mL distilled water+3mL of 10% lead acetate sol.	A white precipitate	[7]
5	Ellagic Acid Test	Plant extract aqueous solution+5% glacial acetic acid+5% sodium nitrite solution	Solution turns muddy / Niger brown precipitate	[3, 16]
6	Potassium dichromate test	Plant extract+few drops of potassium dichromate solution	A dark colour	[27]
7	Hot water test	Warm water in beaker+mature plant part is dipped + warmed for a min.	Black or brown colour ring at the junction of dipping	[34]
8	Test for Carotenoids	(1gm extract+10mL chloroform, vigorously shaken and filtered). Filtrate+conc. H <sub>2</sub> SO <sub>4</sub>	A blue colour at the interface	[35]

**Detection of Tannins:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Gelatin test	Plant extract is dissolved in 5mL distilled water+1% gelatin solution+10% NaCl	A white precipitate	[12, 33]
2	Braymer's test	1mL filtrate <sup>d</sup> +3mL distilled water+3 drops 10% Ferric chloride solution	Blue-green colour	[21, 28]
3	10% NaOH test	0.4mL plant extract+4mL 10% NaOH+shaken well	Formation of emulsion {Hydrolysable tannins}	[21]

4	Bromine water test	10 ml of bromine water + 0.5 gm plant extract	Decoloration of bromine	[23]
5	Lead sub acetate test	1 mL filtrate <sup>e</sup> + 3 drops of lead sub acetate solution	A creamy gelatinous precipitate	[3]
6	Phenazone test	(5 mL aq. extract + 0.5 g of sodium acid phosphate, heated, allowed to cool + filtered); filtrate + 2% solution of phenazone	Precipitation	[9]
7	Mitchell's test	Extract solution + iron + sodium tartarate (+ ammonium acetate solution)	A water-soluble iron-tannin complex, which is insoluble in solution of ammonium acetate	[39]

**Detection of Saponins:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Foam test	0.5 gm plant extract + 2 mL water (vigorously shaken)	Persistent foam for 10 min.	[12]
2	Braymer's test	20 mL water in measuring cylinder + 50 gm extract (vigorously shaken for 15 min.)	Formation of 2 cm thick layer of foam	[7]
3	10% NaOH test	0.2 gm plant extract + 5 mL distilled water; shaken well; heated to boiling	Appearance of creamy mass of small bubbles	[29]
4	NaHCO <sub>3</sub> test	Plant extract + few mL sodium bicarbonate solution + distilled water (vigorously shaken)	Stable honeycomb like froth	[30]

**Detection of Phlobatannins:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	HCl test	2 mL aq. extract + 2 mL 1% HCl (boiled)	A red precipitate	[5, 13]

**Detection of Phytosterols:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Salkowski's test	Filtrate <sup>f</sup> + few drops of conc. H <sub>2</sub> SO <sub>4</sub> (shaken well and allowed to stand)	Red colour (in lower layer)	[21, 12]
2	Liebermann-	50 gm extract is dissolved in 2 mL	An array of colour	[7]



	Burchard's test	acetic anhydride +1-2 drops of conc. H <sub>2</sub> SO <sub>4</sub> (along the side of test tube)	change	
3	Acetic anhydride test	0.5 mL plant extract + 2 mL of acetic anhydride + 2 mL conc. H <sub>2</sub> SO <sub>4</sub>	Change in colour from violet to blue/green	[26]
4	Hesse's response	5 mL aq. extract + 2 mL chloroform + 2 mL conc. H <sub>2</sub> SO <sub>4</sub>	Pink ring/Red colour (in lower chloroform layer)	[23, 40]
5	Sulphur test	Extract solution + pinch of sulphur powder	Sulphur sinks to the bottom	[34]

**Detection of Cholesterol:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1		2 mL extract + 2 mL chloroform + 10 drops of acetic anhydride + 2-3 drops of conc. H <sub>2</sub> SO <sub>4</sub>	A red-rose colour	[22]

**Detection of Terpenoids:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1		2 mL chloroform + 5 mL plant extract, (evaporated on water bath) + 3 mL conc. H <sub>2</sub> SO <sub>4</sub> (boiled on water bath)	A grey coloured solution	[23]

**Detection of Triterpenoids:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Salkowski's test	Filtrate + few drops of conc. H <sub>2</sub> SO <sub>4</sub> (Shaken well and allowed to stand)	Golden yellow layer (at the bottom)	[21]

**Detection of Lignins:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Labat test	Extract solution + gallic acid	An olive green colour	[16, 38]
2	Furfuraldehyde test	Extract solution + 2% furfuraldehyde solution	A red colour	[16, 38]

**Detection of Diterpenes:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Copperacetate test	Plant extract is dissolved in distilled water + 3-4 drops of copper acetate solution	Emerald green colour	[12, 33]

**Detection of Carotenoids:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Carr-Pricer reaction	10 mL extract evaporated to dryness + 2-3 drops of saturated solution of antimony trichloride in chloroform	A blue-green colour eventually changing to red	[22]

**Detection of Anthraquinones:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Borntrager's test	10 mL 10% ammonia sol. + few mL filtrate <sup>g</sup> (shaken vigorously for 30 sec.)	A pink, violet, or red coloured solution	[5, 23, 28]
2	Ammonium hydroxide test	10 mg extract is dissolved in isopropyl alcohol + a drop of conc. ammonium hydroxide solution	Formation of red colour after 2 min.	[32]

**Detection of Anthocyanins:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	HCl test	2 mL plant extract + 2 mL 2N HCl (+ few mL ammonia)	Pink-red sol. which turns blue-violet after addition of ammonia	[18, 31]

**Detection of Leucoanthocyanins:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Isoamyl alcohol test	5 mL plant extract + 5 mL isoamyl alcohol	The upper layer appears red	[31]

**Detection of Carboxylic acid:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Effervescence test	1mL plant extract + 1mL sodium bicarbonate solution	Appearance of effervescence	[21, 26]

**Detection of Coumarins:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	NaOH paper test	0.5 gm moistened extract is taken in a test tube, the mouth of the test tube is covered with 1N NaOH treated filter paper, heated for few min. in water bath	Yellow fluorescence from paper under the UV light	[21, 26]
2	NaOH test	Plant extract + 10% NaOH + Chloroform	A yellow colour	[27]

**Detection of Resins:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Acetic anhydride test	1mL plant extract + Acetic anhydride solution + 1mL conc. H <sub>2</sub> SO <sub>4</sub>	Orange to yellow	[21, 26]
2	Turbidity test	1mL plant extract dissolved in acetone, poured in distilled water	Turbidity	[34]
		10mL extract + 20mL 4% HCl		[36]

**Detection of Emodins:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1		Plant extract + 2mL NH <sub>4</sub> OH + 3mL benzene	A red colour	[29]

**Detection of Gums and Mucilages:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Alcohol test	Dissolve 100mg extract in 10mL distilled water + 25mL absolute alcohol (constant stirring)	White or cloudy precipitate	[7]

**Detection of Fixed oils and Fats:**

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
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1	Spot test/ Stain test	Little quantity of plant extract is pressed in between of filter papers	Oil stain on the paper	[7, 38]
2	Saponification test	Extract + few drops of 0.5N alcoholic KOH + A drop of phenolphthalein (Heated for 2hr.)	Soap formation or partial neutralization of alkali	[7, [38]
3	Saponification test	Extract solution is applied on filter paper	A transparent appearance { oils and resins }	[35, 36]

### Detection of Volatile Oils:

S/N	TEST	PROCEDURE	OBSERVATION (Indicating Positive Test)	REFERENCE
1	Fluorescence test	10mL of extract, filtered till saturation, exposed to UV light	Bright pinkish fluorescence	[37]

<sup>a</sup>=50gm solvent free extract is mixed with few mL dil. HCl and then filtered

<sup>b</sup>=100mg solvent free extract is dissolved in 5mL of distilled water and filtered

<sup>c</sup>=50gm of plant extract is hydrolysed with conc. HCl for 2hr on water bath and filtered

<sup>d</sup>=3gm powdered sample boiled in 50mL distilled water for 3 min. and then filtered

<sup>e</sup>=Small quantity of extract boiled with 5mL of 45% ethanol for 5min. then cooled and filtered

<sup>f</sup>=Equal quantity of chloroform is treated with plant extract and filtered

<sup>g</sup>=3mL of aq. The

extract is shaken with 3mL of benzene and filtered<sup>[5]</sup> OR 10mL of benzene is added in the plant sample and soaked for 10min. then filtered<sup>[23]</sup> OR extract is macerated with ether and filtered<sup>[28]</sup>.

{ }=Indicates the presence of specific phytoconstituents.

**Note:** The composition of various reagents and solutions denoted by italic font have been mentioned in 'Preparation of reagent for Phytochemical Analysis'

### CONCLUSION:

The phytochemical analysis is very much important to evaluate the possible medicinal utilities of a plant and also to determine the active principles responsible for the known biological activities exhibited by the plants. Further, it provides the base for the targeted isolation of compounds and to perform more precise investigations. Extraction of phytochemical from the plant material is mainly

dependent on the type of solvent used. Similarly, the test applied for phytochemical analysis determines the presence or absence of a phytochemical in the sample. Hence, two or more different tests should be performed for more accurate results.

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